SYNTHESIS AND ANTICANDIDAL ACTIVITIES OF OPTIMIZED ANALOGS OF ANTIBIOTIC Sch 37137

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(Received for publication November 24, 1993)

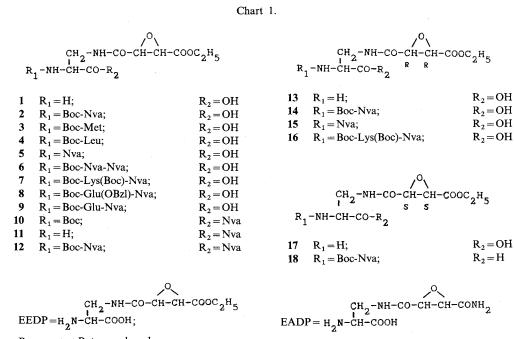
Peptide analogues of Sch 37137 the antifungal antibiotic have been synthesized and evaluated *in vitro* against *Candida* sp. Di- and tripeptides containing methionine, leucine, norvaline, lysine, glutamic acid and N^3 -(*trans*-epoxysuccinamoyl)-L-2,3-diaminopropanoic acid, (EADP) were obtained. Peptides containing (D)-, and (L)-*trans*-epoxysuccinamic acid were also prepared. All of the analogues examined displayed in general higher anticandidal activity than a mixture of diastereomers of Sch 37137.

Fungal infections caused mainly by *Candida albicans* have increased rapidly in the last decades especially in immunocompromised patients such as patients with AIDS, organ transplant recipients, cancer patients undergoing chemotherapy and can cause secondary infections that are life threatening^{1~4)}. Unfortunately, antifungal drugs currently released for clinical use have not offered substantive improvements over amphotericin B, still the drug of choice for most fungal diseases⁵⁾. Therefore, there is still a need for new, safer and more potent antifungal agents.

Fungal cell wall biosynthesis can provide important targets for antifungal agents⁶). Glucosamine-6phosphate synthase, an enzyme which enables the biosynthetic formation of glucosamine containing macromolecules of the fungal cell wall, has been proposed by us as a target for the design of antifungal agents⁷⁾. Our efforts towards rational design of antifungal agents based on this approach have been focused on glucosamine-6-phosphate synthase inhibitors, N³-(4-methoxyfumaroyl)- and N³-iodoacetyl-L-2,3diaminopropanoic acids and their peptide conjugates with antifungal activity^{8~10}). Recently Schering group isolated¹¹⁾ and synthesized¹²⁾ antibiotic Sch 37137, *i.e.* N²-L-alanyl-N³-(D-trans-epoxysuccinamoyl)-L-2,3-diaminopropanoic acid. Structural similarity of this antibiotic to antibiotic A 19009^{13~15} (which differs from Sch 37137 by containing a double bond instead of an epoxide ring) and to N^3 -(4-methoxyfumaroyl)-L-2,3-diaminopropanoic acid (FMDP) derivatives9), both being the glutamine analogs, suggests its mechanism of antifungal action by inhibition of glucosamine-6-phosphate synthase. Our previous report on Sch 37137¹⁶), showed that its diastereoisomeric mixture exhibited a high degree of activity against Candida species. This prompted us to synthesize a series of Sch 37137 analogues in order to improve their in vitro anticandidal activities and to gain information on their structure activity relationships. This report describes the synthesis and in vitro anticandidal evaluation of a series of peptide analogs of Sch 37137.

Chemistry

Classical reactions used in peptide chemistry were employed for the preparation of a series of analogs of Sch 37137, including synthesis of a diastereomeric mixtures as well as both enantiomers. The $2R_{3}R$



configuration of the epoxide moiety of natural Sch 37137 has been established independently by other authors¹²). Most of the syntheses were performed with the use of the racemic trans-epoxysuccinic acid. Our method of synthesis was based in general on the procedure reported earlier for the preparation of Sch 37137 derivatives¹⁶). Compounds obtained during the synthesis are summarized in Chart 1. Treatment of N^3 -(DL-trans-4-ethoxyepoxysuccinyl)-L-2,3-diaminopropanoic acid (EEDP) 1 with Nhydroxy-succinimide esters¹⁷) of Boc-Nva, Boc-Met and Boc-Leu afforded the protected dipeptides 2, 3 and 4, respectively, in good yields. Dipeptide Boc-Nva-EEDP 2 after deptotection of the Boc group was coupled again using the active ester strategy with Boc-Nva, Boc-Lys(Boc) and Boc-Glu(OBzl)¹⁸⁾ to give the protected tripeptides 6, 7 and 8. Boc-EEDP-OSu¹⁶) was reacted with norvaline to give dipeptide 10 which was reacted either with ammonia first followed by deblocking with trifluoroacetic acid to obtain the deprotected dipeptide 10a or deblocked with trifluoroacetic acid first and reacted with active ester of Boc-Nva to furnish the protected tripeptide Boc-Nva-EEDP-Nva 12. Racemic trans-epoxysuccinic acid was resolved into (D)- and (L)-isomers with (D)- or (L)-arginine by diastereomeric salt formation¹⁹⁾. (D)- and (L)-trans-epoxysuccinic acids were converted into their monomethyl esters according to the procedure described earlier¹⁹⁾ and activated with DCC and HOSu to give the active esters which were allowed to react with N^2 -(*tert*-butoxycarbonyl)-L-2.3-diaminopropanoic acid (Boc-A₂pr)²⁰), deblocked and coupled with protected amino acids using the earlier reported methodology to obtain the peptides 14, 16 and 18. The protected peptides after treatment with ammonia followed by acidolysis with trifluoroacetic acid gave the Sch 37137 analogs, $2a \sim 18a$ (Table 1) for antifungal testing.

Results and Discussion

The in vitro activity of Sch 37137 analogs was examined against selected Candida sp. by the broth

Boc = tert-Butoxycarbonyl

No.	Compound	Yield ^a (%)	$[\alpha]_{578}^{25}$ (c 1, MeOH)	MP (°C)	Formula analyses
2a	Nva-EADP	75	+ 9.8		C ₁₂ H ₂₀ N ₄ O ₅ CF ₃ COOH
					Calcd: C 38.88, H 5.32, N 12.95
					Found: C 38.44, H 5.04, N 12.67
3a	Met-EADP	68	+ 6.2	_	C ₁₂ H ₂₀ N ₄ O ₆ SCF ₃ COOH
					Calcd: C 36.36, H 4.57, N 12.11
					Found: C 36.22, H 4.55, N 11.95
4a	Leu-EADP	79	+ 5.8	·	C ₁₃ H ₂₂ N ₄ O ₆ CF ₃ COOH
					Calcd: C 40.54, H 5.21, N 12.60
					Found: C 40.34, H 5.06, N 12.42
6a	Nva-Nva-EADP	72	+ 4.0	—	C ₁₇ H ₂₉ N ₅ O ₇ CF ₃ COOH
					Calcd: C 41.77, H 5.84, N 13.53
					Found: C 41.72, H 5.66, N 13.63
7a	Lys-Nva-EADP	66	- 4.8	—	C ₁₈ H ₃₂ N ₆ O ₇ 2CF ₃ COOH
					Calcd: C 42.92, H 5.56, N 13.65
					Found: C 42.77, H 5.32, N 13.76
9a	Glu-Nva-EADP	81	- 1.2		C ₁₇ H ₂₇ N ₅ O ₉ CF ₃ COOH
					Calcd: C 40.78, H 5.04, N 12.51
					Found: C 40.69, H 4.89, N 12.56
10a	EADP-Nva	70	- 8.0		$C_{12}H_{20}N_4O_6CF_3COOH$
					Caled: C 38.88, H 5.32, N 12.95
					Found: C 38.66, H 5.25, N 12.98
12a	Nva-EADP-Nva	77	+ 8.2	_	C ₁₇ H ₂₉ N ₅ O ₇ CF ₃ COOH
					Calcd: C 41.77, H 5.84, N 13.53
					Found: C 41.52, H 5.82, N 13.31
1 4 a	Nva-EADP ^b	77	-20.2	$120 \sim 122$	C ₁₂ H ₂₀ N ₄ O ₆ CF ₃ COOH
					Calcd: C 38.88, H 5.32, N 12.95
					Found: C 38.68, H 5.29, N 13.05
16a	Lys-Nva-EADP ^b	68	- 9.8		C ₁₈ H ₃₂ N ₆ O ₇ 2CF ₃ COOH
					Calcd: C 42.92, H 5.56, N 13.65
					Found: C 42.84, H 5.41, N 13.66
18a	Nva-EADP ^c	80	+25.8	$138 \sim 140$	C ₁₂ H ₂₀ N ₄ O ₆ CF ₃ COOH
					Calcd: C 38.88, H 5.32, N 12.95
					Found: C 38.77, H 5.19, N 12.79

Table 1. Analytical data of deprotected peptides.

In most cases, trifluoroacetate salts of peptides form hygroscopic, amorphous powders without reproducible melting points.

* Yield of ammonolysis and deprotection.

^b D-Configuration of the epoxy acid.

^c L-Configuration of the epoxy acid.

dilution method in YNB medium²¹⁾ (agar dilution method was also applied for comparison purposes). As shown in Table 2 most of the synthesized peptides appeared to be more active than a racemic mixture of Sch 37137 previously prepared in our laboratory¹⁶⁾. However, among the peptides obtained, the activity changed depending on the amino acid incorporated into the peptide chain. Dipeptide with norvaline residue in the *N*-terminal position was shown to be the most active peptide tested. Also lengthening of a peptide chain altered the anticandidal activity. In general, tripeptides were somewhat less potent than dipeptides. Chirality of the *trans*-epoxysuccinic acid was another factor influencing on the activity. Thus, dipeptide having D-*trans*-epoxysuccinamic acid (2*R*,3*R* isomer) showed higher activity than compound with the L- acid (approximately 20 times higher) suggesting that the D chirality is essential for antifungal activity. Accordingly, the tripeptide Lys-Nva-EADP with the (2*R*,3*R*) configuration of the epoxy acid exhibited higher activity than corresponding peptide with a racemic epoxy acid. Interestingly, peptide 11

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	MIC^{a} ($\mu g/ml$)						
Compound	Candida albicans ATCC 26278 YNB medium		Candida krusei	Candida glabrata	Candida famata 1940	Candida albicans	
	Broth	Agar		0		2043	
Ala-EADP ^b	1.2	2.0	NT	1.5	NT	1.0	
Nva-EADP	0.25	0.75	0.35	0.75	0.50	0.25	
Met-EADP	0.35	0.75	1.0	1.25	0.5	0.5	
Leu-EADP	0.3	0.75	0.5	0.75	0.5	0.25	
Nva-Nva-EADP	0.75	2.0	1.2	1.0	1.0	0.75	
Lys-Nva-EADP	0.50	1.0	1.5	0.75	1.0	0.50	
Glu-Nva-EADP	0.75	1.5	1.2	1.5	0.75	0.50	
EADP-Nva	0.5	1.5	1.0	1.25	0.75	0.75	
Nva-EADP-Nva	0.75	2.0	1.5	1.5	1.0	0.50	
Nva-EADP°	0.15	0.50	0.35	0.5	0.35	0.10	
Lys-Nva-EADP°	0.20	0.50	0.75	0.50	0.50	0.25	
Nva-EADP ^d	7.5	15.0	10.0	12.5	10.0	6.50	
EEDP-Nva	12.5	25.0	>15.0	>15.0	>15.0	>15.0	
Lys-Nva-FMDP	0.75	1.5	1.0	1.5	0.75	0.7	

Table 2. In vitro activity of Sch 37137 analogues.

Peptides were tested as trifluoroacetate salts.

^a Yeast nitrogen base (Difco) - YNB medium containing 200 μg/ml of sodium glutamate, 10⁶ cfu/ml, 30°C, 24 hours; agar method (YNB medium with 2% agar).

^b Diasteromeric mixture of Sch 37137.

^c D-Configuration of the epoxy acid.

^d L-Configuration of the epoxy acid.

Table 3. MIC, transport and intracellular cleavage rates of selected EADP-peptides in *Candida albicans* ATCC 26278.

Compound	MIC (µg/ml)	Transport rate (nmol/minute /mg protein)	Intracellular cleavage rate (nmol/minute /mg protein)
Nva-EADP	0.25	2.0	20.4
Leu-EADP	0.3	2.3	27.0
Lys-Nva-EADP*	0.2	4.2	37.5
Nva-Nva-EADP	0.75	1.4	16.3
Nva-EADP-Nva	0.75	2.4	4.9

^a D-Configuration of the epoxy acid.

Table 4. Change of antifungal activity of EADPpeptides against *Candida albicans* ATCC 26278 in serum protein solution^a expressed as % of standard activity.

Compound	5% HSA ^b	HS⁵
Leu-EADP	100	25
Nva-EADP	100	10
Lys-Nva-EADP	85	20
Nva-Nva-EADP	75	25
Nva-EADP-Nva	60	30

Growth was performed in the 5% solution of serum albumin in YNB modified medium or 1:1 mixture of YNB modified medium and serum, but peptides were previously preincubated in the presence of serum proteins for 2 hours at 37°C before inoculation.

^b HSA—Human serum albumin, HS—Human serum.

(EEDP-Nva) containing an ethyl ester instead of an amide group in the *trans*-epoxy acid moiety showed weak anticandidal activity. The most active peptide containing N^3 -(4-methoxyfumaroyl)-L-2,3-diaminopropanoic acid (FMDP), *i.e.* Lys-Nva-FMDP⁹⁾ was also used for comparative testing. This compound however was less active than the Sch 37137 analogues obtained in this study.

The uptake of various peptides with the racemic epoxide moiety (Table 3) into fungal cells showed approximately the same range of velocities. Moreover the rate of intracellular cleavage by peptidases ranged from 4.9 (for Nva-EADP-Nva) to a high value of 37.5 nmol/minute/mg protein (for Lys-Nva-EADP) and showed good correlation with antifungal activity. We have also shown that the peptides were partially

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inactivated by human serum components, thus affecting the effectiveness of the Sch 37137 and its analogs against fungal cells. The same phenomenon was observed earlier by us for N^3 -(4-methoxyfumaroyl)-L-2,3-diaminopropanoic acid peptides²²⁾. Incubation of the peptides (Table 4) with 5% human serum albumin and especially in complete human resulted in the significant loss of their antifungal efficacy. It suggested the interaction of serum proteins with the peptides by an unknown mechanism which needs further investigation.

In conclusion, a series of Sch 37137 analogs have been synthesized and evaluated for *in vitro* anticandidal activity. These peptides are taken up into cells *via* peptide permeases ("portage" transport²³⁾), followed by enzymatic hydrolysis inside the cell and intracellular generation of the enzyme inhibitor. Anticandidal activity is influenced by the length of the peptide chain, the kind of amino acid incorporated into the peptide and the chirality of the *trans*-epoxysuccinic acid.

Experimental

¹H NMR spectra were recorded at 60 MHz on a Varian 360 instrument with Me₄Si as internal reference. Optical rotations were measured in a Polamat A (Carl Zeiss Jena) polarimeter. Melting points were determined in an open capillary tubes and are uncorrected. TLC was carried out using Kieselgel 60 (Merck) plates. The analytical results obtained for C, H and N were within $\pm 0.4\%$ of the theoretical values.

The *N*-hydroxysuccinimide esters of *N*-(*tert*-butoxycarbonyl)amino acids (Nva, Lys, Met, Leu and Glu(OBzl)) were prepared by the previously described method¹⁷. Monoethyl L- and D-*trans*-epoxysuccinate were prepared according to the published procedure¹⁹.

Representative synthetic route to a Sch 37137 analogue.

 $\frac{N^2-(tert-Butoxycarbonyl-L-norvalyl)-N^3-(DL-trans-4-ethoxyepoxysuccinyl)-L-2,3-diaminopropanoic}{2 (Boc-Nva-EEDP)}$

To a cooled solution of N^3 -(DL-*trans*-4-ethoxyepoxysuccinyl)-L-2,3-diaminopropanoic acid trifluoroacetate salt¹⁶ 1 (1.44 g, 4 mmol) and NaHCO₃ (0.69 g, 8 mmol) in a mixture of water - methanol (1 : 1 v/v) (10 ml) was added Boc-Nva-OSu (1.25 g, 4 mmol) in MeOH (10 ml) with stirring. The reaction mixture was stirred for 1 hour in an ice-bath then at room temperature overnight. After the usual work-up and evaporation of the solvents compound **2** (1.69 g, 90% yield) was obtained. MP 69~70°C, $[\alpha]_{578}^{25}$ -14.2° (c 1, MeOH), ¹H NMR (DMSO-d₆) δ 0.9 (3H, m), 1.1 (3H, t, J=7 Hz), 1.2 (9H, s), 1.3~1.5 (4H, m), 3.5~3.7 (4H, m), 3.8 (1H, m), 4.1 (1H, m), 4.3 (2H, q, J=7 Hz), 6.6~6.8 (2H, m), 7.2 (1H, m).

Anal Calcd for $C_{19}H_{31}N_3O_9$:C 51.22, H 7.01, N 9.43.Found:C 51.38, H 7.27, N 9.21.

The following compounds were also prepared.

$\frac{N^2-(tert-Butoxycarbonyl-L-methionyl)-N^3-(DL-trans-4-ethoxyepoxysuccinyl)-L-2,3-diaminopro$ panoic Acid 3

1.46 g (77% yield). MP 81~83°C, $[\alpha]_{578}^{25}$ -12.8° (c 1, MeOH), ¹H NMR (DMSO-d₆) δ 1.1 (3H, t, J=7 Hz), 1.1~1.3 (2H, m), 1.4 (9H, s), 2.1 (3H, s), 2.2~2.4 (2H, m), 2.6 (2H, m), 3.5~3.7 (4H, m), 3.8 (1H, m), 4.2 (1H, m), 6.7 (2H, m), 7.2 (1H, m).

 Anal Calcd for C₁₉H₃₁N₃O₉S:
 C 47.78, H 6.54, N 8.79.

 Found:
 C 47.44, H 6.62, N 8.59.

 $\frac{N^2-(tert-Butoxycarbonyl-L-leucyl)-N^3-(DL-trans-4-ethoxyepoxysuccinyl)-L-2,3-diaminopropanoic}{4}$

1.5 g (82% yield). MP 79~80°C, $[\alpha]_{578}^{25}$ -26.2° (c 1, MeOH), ¹H NMR (DMSO-d₆) δ 0.9~1.4 (21H, br m), 1.5 (2H, m), 3.4~3.7 (4H, m), 3.8 (1H, m), 4.1 (1H, m), 4.3 (2H, q, J=7 Hz), 6.6~6.8 (2H, m),

7.2 (1H, m).

Anal Calcd for C₂₀H₃₃N₃O₉: C 52.26, H 7.23, N 9.15. Found: C 52.04, H 7.26, N 9.22.

 $\frac{N^2-L-Norvalyl-N^3-(DL-trans-4-ethoxyepoxysuccinyl)-L-2,3-diaminopropanoic Acid Trifluoroacetate}{5}$

Compound 2 (1.33 g, 3 mmol) was dissolved in cold trifluoroacetic acid and kept at room temperature for 2 hours. The solvent was removed *in vacuo*, the residue triturated with dry ethyl ether and dried over KOH to give 1.3 g (95% yield) of **5** as an amorphous hygroscopic trifluoroacetic acid salt. $[\alpha]_{578}^{25} - 5.2^{\circ}$ (*c* 1, MeOH), ¹H NMR (D₂O) δ 1.0 (3H, m), 1.2 (3H, t, J=7 Hz), 1.3~1.5 (4H, m), 3.5~3.6 (2H, m), 3.7 (2H, m), 3.8 (1H, m), 4.0 (2H, q, J=7 Hz), 4.1 (1H, m).

Anal Calcd for C₁₄H₂₃N₃O₇CF₃COOH: C 41.83, H 5.26, N 9.14. Found: C 41.66, H 5.14, N 9.02.

 N^2 -[(tert-Butoxycarbonyl-L-norvalyl)-L-norvalyl]- N^3 -(DL-trans-4-ethoxyepoxysuccinyl)-L-2,3-diaminopropanoic Acid **6**

This compound was prepared from Boc-Nva-OSu and compound 5 according to the procedure described for the preparation of 2. 0.47 g (83% yield). MP 87~90°C, $[\alpha]_{578}^{25}$ -38.0° (c 1, MeOH).

Anal Calcd for C₂₄H₄₀N₄O₁₀: C 52.92, H 7.40, N 10.28. Found: C 52.71, H 7.22, N 10.06.

 $\frac{N^2 - [N^{\alpha}, N^{\varepsilon} - \text{Bis}(tert-butoxycarbonyl-L-lysyl)-L-norvalyl] - N^3 - (DL-trans-4-ethoxyepoxysuccinyl)-L-2, 3-diaminopropanoic Acid 7$

According to the methodology described for 2, peptide 7 was prepared, 0.54 g (80%). MP 62~65°C, $[\alpha]_{578}^{25} - 25.8^{\circ}$ (c 1, MeOH).

Anal Calcd for C₃₀H₅₁N₅O₁₂: C 53.47, H 7.63, N 10.39. Found: C 53.21, H 7.56, N 10.42.

 N^2 -[(tert-Butoxycarbonyl- γ -O-benzyl-L-glutamyl)-L-norvalyl]- N^3 -(DL-trans-4-ethoxyepoxysuccinyl)-L-2,3-diaminopropanoic Acid 8

Tripeptide 8 was obtained from Boc-Glu(OBzl)OSu and peptide 5 according to the method described for 2. 0.59 g (83%). MP 68~70°C, $[\alpha]_{578}^{25}$ -24.0° (c 1, MeOH).

Anal Calcd for $C_{31}H_{44}N_4O_{12}$:C 55.92, H 6.67, N 8.42.Found:C 55.66, H 6.63, N 8.54.

 $\frac{N^2-[(tert-Butoxycarbonyl-L-glutamyl)-L-norvalyl]-N^3-(DL-trans-4-ethoxyepoxysuccinyl)-L-2,3-di-aminopropanoic Acid 9$

Peptide 8 0.58 g (0.9 mmol) was dissolved in 20 ml MeOH and hydrogenolyzed in the presence of 0.05 g of 10% palladium catalyst on carbon. Then the catalyst was filtered off, the filtrate was concentrated and the residue crystallized from methanol-ethyl ether-hexane mixture to give 0.48 g (95%) of 9. MP $92 \sim 94^{\circ}$ C, $[\alpha]_{578}^{25} - 32.0^{\circ}$ (c 1, MeOH).

Anal Calcd for C₂₄H₃₈N₄O₁₂: C 50.16, H 6.66, N 9.75. Found: C 50.08, H 6.68, N 9.44.

 $(N^2-(tert-Butoxycarbonyl)-N^3-[(DL-trans-4-ethoxyepoxysuccinyl)-L-2,3-diaminopropanoyl])-L$ norvaline 10

From N^2 -(*tert*-butoxycarbonyl)- N^3 -(DL-*trans*-4-ethoxyepoxysuccinyl)-L-2,3-diaminopropanoic acid succinimido ester¹⁶) and L-norvaline by the procedure used to prepare 2, compound 10 was obtained. 1.12 g (75%). MP 72~74°C, $[\alpha]_{578}^{25} - 14.0^{\circ}$ (c 1, MeOH).

Anal Calcd for $C_{19}H_{31}N_3O_9$:C 51.22, H 7.01, N 9.43.Found :C 51.37, H 6.82, N 9.66.

$(N^2-(tert-Butoxycarbonyl-L-norvalyl)-N^3-[(DL-trans-4-ethoxyepoxysuccinyl)-L-2,3-diaminopro-$
panoyl])-L-norvaline 12
12 was prepared similarly to 2 from freshly deprotected peptide 11 (after trifluoroacetic acid treatment
of 10) and Boc-Nva-OSu. 0.41 g (71%). MP $103 \sim 107^{\circ}$ C, $[\alpha]_{578}^{25} - 30.2^{\circ}$ (c 1, MeOH).
Anal Calcd for $C_{24}H_{40}N_4O_{10}$: C 52.92, H 7.40, N 10.28.
Found : C 52.77, H 7.34, N 10.16.
N ³ -(p-trans-4-Ethoxyepoxysuccinyl)-L-2,3-diaminopropanolic Acid Trifluoroacetate Salt 13
N-Succinimido ethyl D- <i>trans</i> -epoxysuccinate [MP 96~98°C, $[\alpha]_{578}^{25}$ -70.4° (c 1, THF), ¹ H NMR
$(DMSO-d_6)$; δ 1.3 (3H, t, $J=7$ Hz), 2.9 (4H, s), 3.85 (1H, d, $J=2$ Hz), 3.95 (1H, d, $J=2$ Hz), 4.35 (2H,
q, $J = 7$ Hz). Anal Calcd for C ₁₀ H ₁₁ NO ₂ : C 46.69, H 4.31, N 5.44. Found: C 46.58, H 4.40, N 5.51.]
was prepared according to the procedure described for the preparation of the racemic compound ¹⁶). This
was reacted with N^2 -(<i>tert</i> -butoxycarbonyl)-L-2,3-diaminopropanoic acid, following the procedure published
for the preparation of N^3 -(DL-trans-4-ethoxyepoxysuccinyl)-L-2,3-diaminopropanoic acid ¹⁶), yielding 0.31 g
(96%) of compound 13. MP 164~166°C, $[\alpha]_{578}^{25} - 71.2^{\circ}$ (c 1, H ₂ O).
Anal Calcd for C ₉ H ₁₄ N ₂ O ₆ CF ₃ COOH: C 36.67, H 4.19, N 7.77.
Found : C 36.30, H 4.05, N 7.42.
N^2 (and Determine the analysis of the second state of the superscription $\lambda = 2.2$ diamin encourses
N^2 -(<i>tert</i> -Butoxycarbonyl-L-norvalyl)- N^3 -(D- <i>trans</i> -4-ethoxyepoxysuccinyl)-L-2,3-diaminopropanoic
$\frac{\text{Acid } 14}{14}$ was prepared from Boc-Nva-OSu and 13 as described for the preparation of 2 in 85% yield. MP
$76 \sim 78^{\circ}$ C, $[\alpha]_{278}^{25} - 49.8^{\circ}$ (c 1, MeOH).
Anal Calcd for $C_{19}H_{31}N_3O_9$: C 51.22, H 7.01, N 9.43.
Found: $C 51.12, H 7.12, N 9.38.$
N^2 -L-Norvalyl- N^3 -(D- <i>trans</i> -4-ethoxyepoxysuccinyl)-L-2,3-diaminopropanoic Acid Trifluoroacetate
Salt 15 \overline{E} and 1.1 σ (2.5 mm c)) of 15 mm c) to 14 of the trifference of the product of 15 mm c) to 15 mm
From 1.1 g (2.5 mmol) of peptide 14 after trifluoroacetic acid treatment 0.93 g (90%) of 15 was obtained.
MP $102 \sim 106^{\circ}$ C, $[\alpha]_{578}^{25} - 26.2^{\circ}$ (c 1, MeOH). Anal Calcd for C ₁₄ H ₂₃ N ₃ O ₇ CF ₃ COOH: C 41.83, H 5.26, N 9.14.
Found: $C 41.51, H 5.12, N 9.03.$
Found: C 41.51, H 5.12, N 5.05.
N^2 -[$N^{\alpha}, N^{\varepsilon}$ -Bis(<i>tert</i> -butoxycarbonyl-L-lysyl)-L-norvalyl]- N^3 -(D- <i>trans</i> -4-ethoxyepoxysuccinyl)-L-2,3-
diaminopropanoic Acid 16
This compound was prepared from Boc-Lys(Boc)-OSu and 15 by the method used for the preparation
of the racemic 7, 1.16 g (82%). MP 79 ~ 81°C, $[\alpha]_{578}^{25}$ -47.2° (c 1, MeOH).
Anal Calcd for $C_{30}H_{51}N_5O_{12}$: C 53.47, H 7.63, N 10.39.
Found: C 53.34, H 7.44, N 10.44.
$\frac{N^3-(L-trans-4-Ethoxyepoxysuccinyl)-L-2,3-diaminopropanoic Acid Trifluoroacetate Salt 17}{17}$
17 was obtained analogously to the preparation of 13 using N-succinimido ethyl L-trans-epoxysuc-
17 was obtained analogously to the preparation of 13 using N-succinimido ethyl L-trans-epoxysuc- cinate [MP 97.5~98.5°C, $[\alpha]_{578}^{25}$ + 68.8° (c 1, THF) ¹ H NMR (DMSO-d ₆) δ 1.25 (3H, t, J=7 Hz), 2.9
17 was obtained analogously to the preparation of 13 using N-succinimido ethyl L-trans-epoxysuc- cinate [MP 97.5~98.5°C, $[\alpha]_{578}^{25}$ + 68.8° (c 1, THF) ¹ H NMR (DMSO-d ₆) δ 1.25 (3H, t, J=7Hz), 2.9 (4H, s), 3.8 (1H, d, J=2Hz), 3.9 (1H, d, J=2Hz), 4.35 (2H, q, J=7Hz). Anal Calcd for C ₁₀ H ₁₁ NO ₇ :
17 was obtained analogously to the preparation of 13 using N-succinimido ethyl L-trans-epoxysuc- cinate [MP 97.5~98.5°C, $[\alpha]_{578}^{25}$ + 68.8° (c 1, THF) ¹ H NMR (DMSO-d ₆) δ 1.25 (3H, t, J=7 Hz), 2.9

Anal Calcd for C₉H₁₄N₂O₆CF₃COOH: C 36.67, H 4.19, N 7.77. Found: C 36.41, H 3.98, N 7.54.

 $\frac{N^2 - (tert-Butoxycarbonyl-L-norvalyl) - N^3 - (L-trans-4-ethoxyepoxysuccinyl) - L-2, 3-diaminopropanoic}{18}$

succinyl)-L-2,3-diaminopropanoic acid¹⁶). 0.35 g (95%). MP 150~152°C and $[\alpha]_{578}^{25}$ +29.8° (c 1, H₂O).

18 was prepared similarly to **14** from Boc-Nva-OSu and **17** with 78% yield. $[\alpha]_{578}^{25}$ + 30.2° (c, MeOH).

General Procedure for Ammonolysis and Deprotection of Peptides

The appropriate peptide 2, 3, 4, 6, 7, 9, 10, 12, 14, 16 and 18 (1 mmol) was stirred in a cold ammonia solution (29%), 20 ml for 2 hours, then ammonia was evaporated, the crude residue dissolved in 5 ml of water and passed through a short column of Amberlite CG 50 (H^+), washed with water, evaporated and dried over KOH, then treated with 5 ml cold trifluoroacetic acid for 2 hours. The solvent was removed under reduced pressure leaving the deprotected peptides 2a, 3a, 4a, 6a, 7a, 9a, 10a, 12a, 14a, 16a and 18a. Yields and analytical data are collected in Table 1.

Acknowledgments

We would like to thank The State Committee for Scientific Research Warsaw for financial support of these studies.

References

- MEUNIER, F.; A. LELEUX, R. SNOECK, J. GERAIN, C. LAMBERT & A. M. CEUPPENS: Chemoprophylaxis of fungal infections. *In* Diagnosis and Therapy of Systemic Fungal Infections. *Eds.*, K. HOLMBERG & R. MEYER, pp. 167~177, Raven Press, New York, 1989
- PERFECT, J. F.; W. W. PICKARD, D. L. HUNT, B. PALMER & W. A. SCHELL: The use of amphotericin B in nosocomial fungal infection. Rev. Infect. Dis. 13: 474~479, 1991
- 3) SARAL, R.: Candida and Aspergillus infections in immunocompromised patients: an overview. Rev. Infect. Dis. 13: 487~492, 1991
- WALSH, T. J. & A. PIZZO: Treatment of systemic fungal infections: Recent progress and current problems. Eur. J. Clin. Microbiol. Infect. Dis. 7: 460~475, 1988
- GALLIS, H. A.; R. DREW & W. W. PICKARD: Amphotericin B: 30 years of clinical experience. Rev. Infect. Dis. 12: 308 ~ 329, 1990
- CASSONE, A.: Cell walls of pathogenic yeasts and implications for antimycotic therapy. Drugs Exp. Clin. Res. 12: 635~643, 1986
- 7) BOROWSKI, E.: Targets for antifungal drugs: a critical overview. Abstract of the 8th International Symposium on Future Trends in Chemotherapy, p. 158, Tirrenia (Italy), Mar. 28~30, 1988
- ANDRUSZKIEWICZ, R.; H. CHMARA, S. MILEWSKI & E. BOROWSKI: Synthesis of N³-fumaramoyl-L-2,3-diaminopropanoic acid analogues, the irreversible inhibitors of glucosamine synthetase. Int. J. Peptide Protein Res. 27: 449~453, 1986
- ANDRUSZKIEWICZ, R.; S. MILEWSKI, T. ZIENIAWA & E. BOROWSKI: Anticandidal properties of N³-(4-methoxy-fumaroyl)-L-2,3-diaminopropanoic acid oligopeptides. J. Med. Chem. 33: 132~135, 1990
- ANDRUSZKIEWICZ, R.; H. CHMARA, S. MILEWSKI, T. ZIENIAWA & E. BOROWSKI: Antimicrobial properties of N³-(iodoacetyl)-L-2,3-diaminopropanoic acid—peptide conjugates. J. Med. Chem. 33: 2755~2759, 1990
- 11) COOPER, R.; A. C. HORAN, F. GENTILE, V. GULLO, D. LOEBENBERG, J. MARQUEZ, M. PATEL, M. S. PUAR & I. TRUUMEES: Sch 37137, a novel antifungal compound produced by a *Micromonospora* sp. Taxonomy, fermentation, isolation, structure and biological properties. J. Antibiotics 41: 13~19, 1988
- 12) RANE, D. F.; V. M. GIRIJAVALLABHAN, A. K. GANGULY, R. E. PIKE, A. K. SAKSENA & A. T. MCPHAIL: Total synthesis and absolute stereochemistry of the antifungal dipeptide Sch 37137 and its 2S,3S isomer. Tetrahedron Lett. 34: 3201~3204, 1993
- 13) MOLLOY, B. B.; D. H. LIVELY, R. M. GALE, M. GORMAN, L. D. BOECK, C. E. HIGGENS, R. E. KASTNER, L. L. HUCKSTEP & N. NEUSS: A new dipeptide antibiotic from *Streptomyces collinus*, Lindenbein, J. Antibiotics 25: 137~140, 1972
- 14) VAN DER BAAN, J. L.; J. W. F. K. BARNICK & F. BICKELHAUPT: Antibiotic A 19009. Structural investigation and synthesis. J. Antibiotics 36: 784~792, 1983
- 15) ANDRUSZKIEWICZ, R.; H. CHMARA & E. BOROWSKI: Anticandidal activity of antibiotic A 19009 and its isomer. J. Antibiotics 37: 1479~1482, 1984
- ANDRUSZKIEWICZ, R.: Synthesis and antifungal properties of diastereomers and analogs of antibiotic Sch 37137.
 J. Antibiotics 47: 380 ~ 385, 1994
- 17) ANDERSON, G. W.; J. E. ZIMMERMAN & F. M. CALLAHAN: The use of esters of N-hydroxysuccinimide in peptide synthesis. J. Am. Chem. Soc. 86: 1839~1842, 1964
- 18) NAKAJIMA, K. & K. OKAWA: Synthesis of the synthetic model enzyme. The synthesis of Cyclo(His-Glu-Cys-D-

Phe-Gly)₂ as the esterase model. Bull. Chem. Soc. Jpn. 46: $1811 \sim 1816$, 1973

- 19) TAMAI, M.; C. YOKOO, M. MURATA, K. OGUMA, K. SOTA, E. SATO & Y. KANAOKA: Efficient synthetic method for ethyl (+)-(2S,3S)-3-[(S)-3-methyl-1-(3-methylbutylcarbamoyl)butylcarbamoyl]-2-oxiranecarboxylate (EST), a new inhibitor of cysteine proteinases. Chem. Pharm. Bull. 35: 1098~1104, 1987
- 20) WAKI, M.; Y. KITAJIMA & N. IZUMIYA: A facile synthesis of N²-protected L-2,3-diaminopropanoic acid. Synthesis 1981: 266~268, 1981
- 21) MILEWSKI, S.; H. CHMARA, R. ANDRUSZKIEWICZ, E. BOROWSKI, M. ZAREMBA & J. BOROWSKI: Antifungal peptides with novel specific inhibitors of glucosamine-6-phosphate synthase. Drugs Exp. Clin. Res. 14: 461~465, 1988
- 22) KASPRZAK, L.; S. MILEWSKI, J. GUMIENIAK & E. BOROWSKI: The influence of serum proteins on biological activity of anticandidal peptides containing N^3 -(4-methoxyfumaroyl)-L-2,3-diaminopropanoic acid. J. Chemother. 4: 88~94, 1992
- 23) FICKEL, T. E. & C. GILVARG: Transport of impermeant substances in *E. coli* by way of the oligopeptide permease. Nature (London) 241: 161~163, 1973